

Update on Clinical Topics in Antiretroviral Therapy workshop

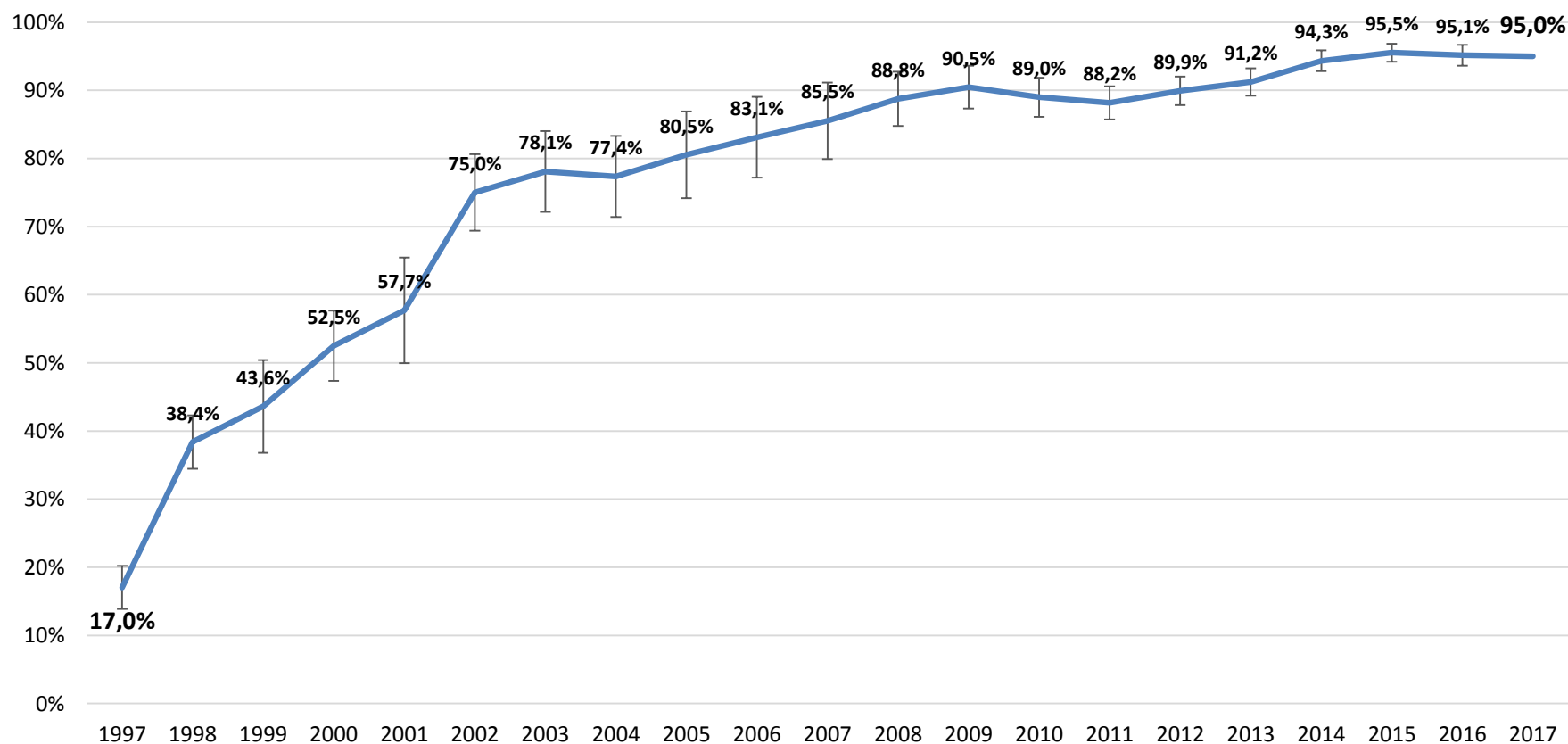
Hospital de Sant Pau Art Nouveau Site, Barcelona, Spain

Friday 27th and Saturday 28th April 2018

Implications from the virology field in successful ART

Carlo-Federico Perno

Proportion of patients with a VL \leq 80 copies/mL at 12 months from starting their first ART regimen by calendar year of initiation



In the long-term management of HIV infection, the stable maintenance of undetectable HIV over the years is more important than the mere success of the first regimen

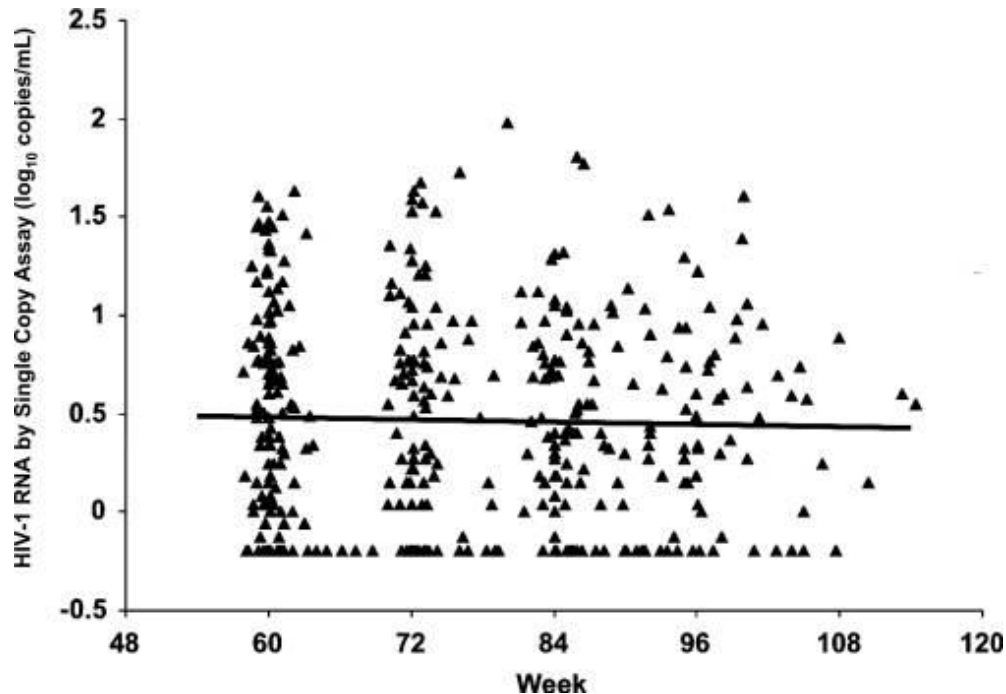
Definition of virologically suppressed

Clinical trials exploring switching strategies have defined suppression as an HIV-VL < 50 copies/mL for at least 6 months.

- Routine commercial assays can now accurately measure viraemia levels as low as 50 copies/mL and as high as 10 million or more copies/mL.
- Among them, the two most widely available real-time reverse transcription PCR assays have reported RNA quantification limits of 40 copies/mL (Abbot RealTime RT-PCR Assay) and 20 copies/mL (Roche COBAS Amplicor TaqMan assay version 2.0), and can qualitatively detect HIV-1 RNA below these quantification limits.
- **More sensitive research assays have reduced the lower end to much less than one copy/mL with good quantitative accuracy.**

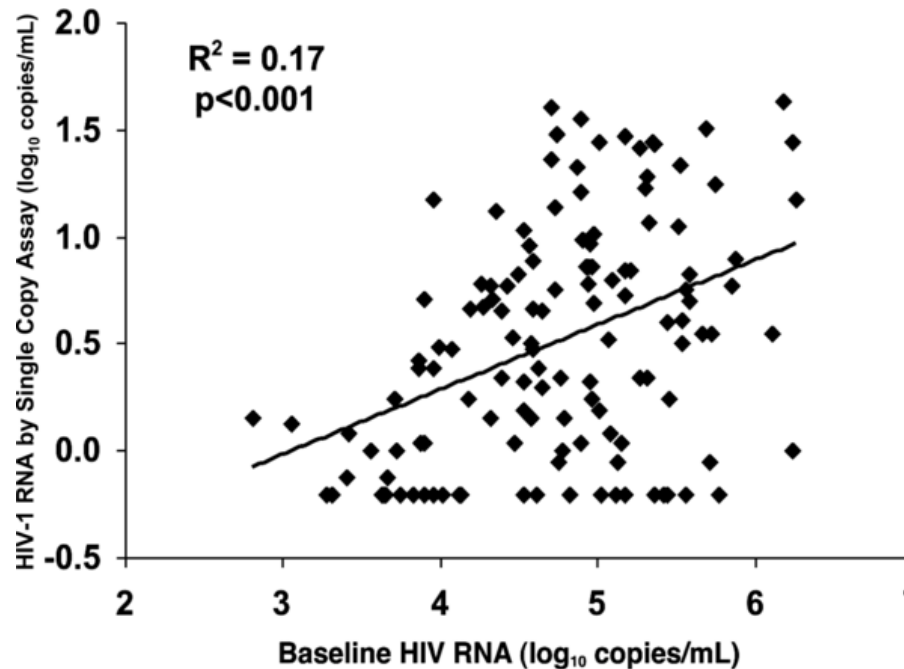
Single copy assay results revealed that >80% of patients on initial antiretroviral therapy for 60 wks had persistent viremia of one copy/ml or more with an overall median of 3.1 (range 1-49) copies/ml.

HIV-1 RNA Levels Over 50 wks of Suppressive Antiretroviral Therapy

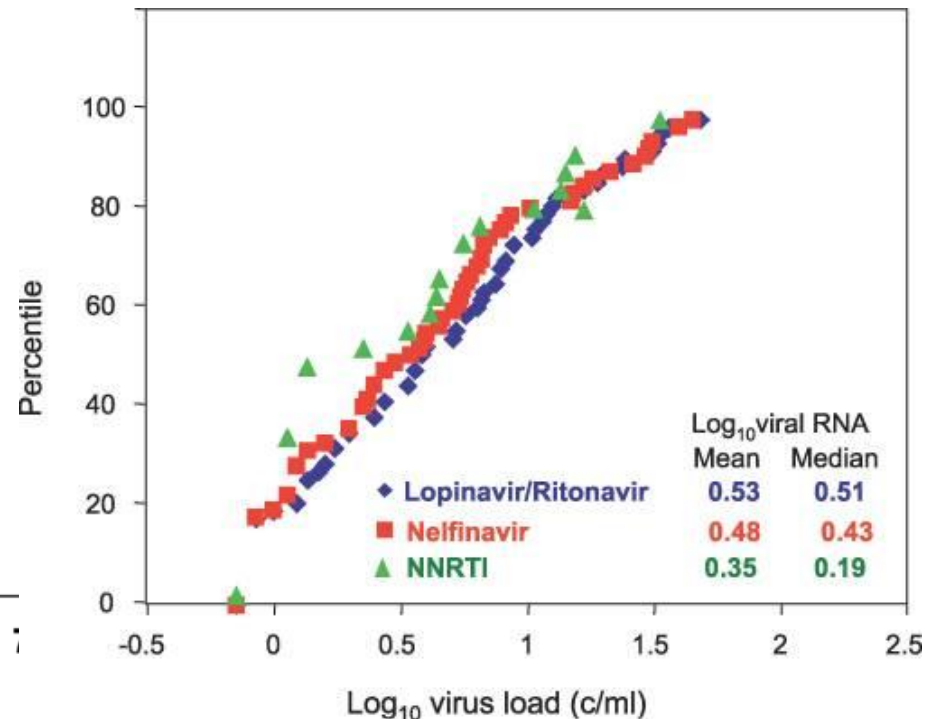


Longitudinal studies revealed no significant decline in the level of viremia between 60 and 110 wks of suppressive antiretroviral therapy. These data suggest that the persistent viremia on current antiretroviral therapy is derived, at least in part, from long-lived cells that are infected prior to initiation of therapy.

The level of viremia correlated with pretherapy plasma HIV-1 RNA but not with specific treatment.



Correlation between Pretherapy and On-Therapy Viral RNA Levels



Distribution of Plasma HIV-1 RNA Levels in Patients with Persistently Suppressed Viremia (HIV-1 RNA <50 Copies/ml from Week 24 to 60) on Standard Antiretroviral Therapy

Several studies demonstrated that an increased risk of virological failure remains even in responder patients with very low copies of viral replication during ART.

- Lambert-Niclot S, et al. Analysis and impact of ultra sensitive viral load on virological failure in 3 protease inhibitor monotherapy trials as simplification regimen (ANRS and IMEA Studies). IAS 2017, abstract MOPEB0318.
- Gianotti N, et al., Refining criteria for selecting candidates for a safe lopinavir/ritonavir or darunavir/ritonavir monotherapy in HIV-infected virologically suppressed patients. PLoS One 2017;12(2):e0171611.
- Gianotti et al., HIV DNA loads, plasma residual viraemia and risk of virological rebound in heavily treated, virologically suppressed HIV-infected patients. Clin Microbiol Infect. 2015 21:103.e7-103.e10.
- Álvarez Estévez M, et al. Quantification of viral loads lower than 50 copies per milliliter by use of the Cobas AmpliPrep/Cobas TaqMan HIV-1 test, version 2.0, can predict the likelihood of subsequent virological rebound to >50 copies per milliliter. J Clin Microbiol. 2013, 51:1555-7.
- Maggiolo F, et al. Ultrasensitive assessment of residual low-level HIV viremia in HAART-treated patients and risk of virological failure. JAIDS. 2012;60:473-82.
- Doyle T, et al.. Plasma HIV-1 RNA detection below 50 copies/ml and risk of virologic rebound in patients receiving highly active antiretroviral therapy. CID 2012, 54:724-32.
- Charpentier C, et al. Persistent low-level HIV-1 RNA between 20 and 50 copies/mL in antiretroviral-treated patients: associated factors and virological outcome. JAC 2012.67:2231-5.
- Lambert-Niclot S, et al. Factors associated with virological failure in HIV-1-infected patients receiving darunavir/ritonavir monotherapy. J Infect Dis. 2011, 204:1211-6.
- Bonora S, et al. 2009. Ultrasensitive assessment of residual HIV viremia in HAART-treated patients with persistently undetectable plasma HIV-RNA: a cross-sectional study. J Med Virol. 81:400–405.

Factors influencing long-term viral suppression

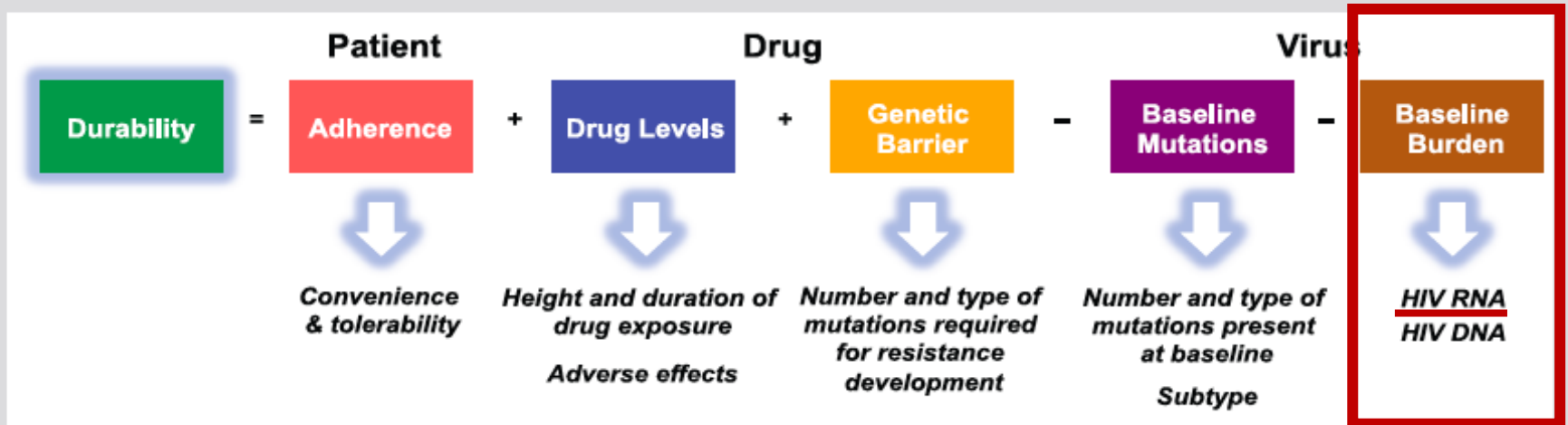


Figure 1. Factors influencing long-term viral suppression, adapted from Massimo Andreoni personal communication.

Pre-HAART viremia has an impact on the achievement of virological success.....

By 72 weeks of therapy, patients having **pre-HAART viremia >500,000 copies/mL** showed the lowest probability of achieving VS compared to others pre-HAART viremia ranges.

Pre-HAART viremia ranges
(copies/mL):

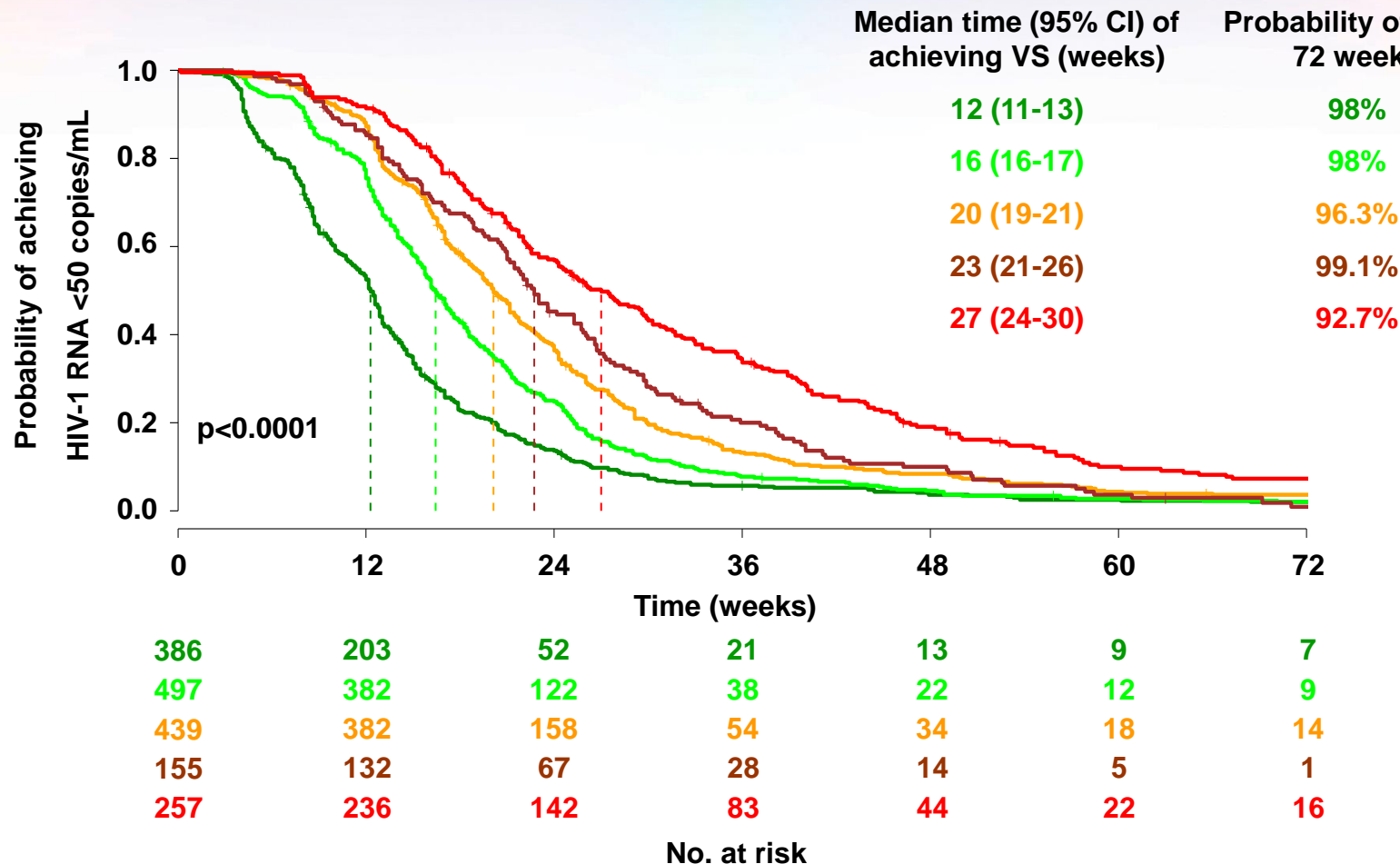
<30K

30-100K

100-300K

300-500K

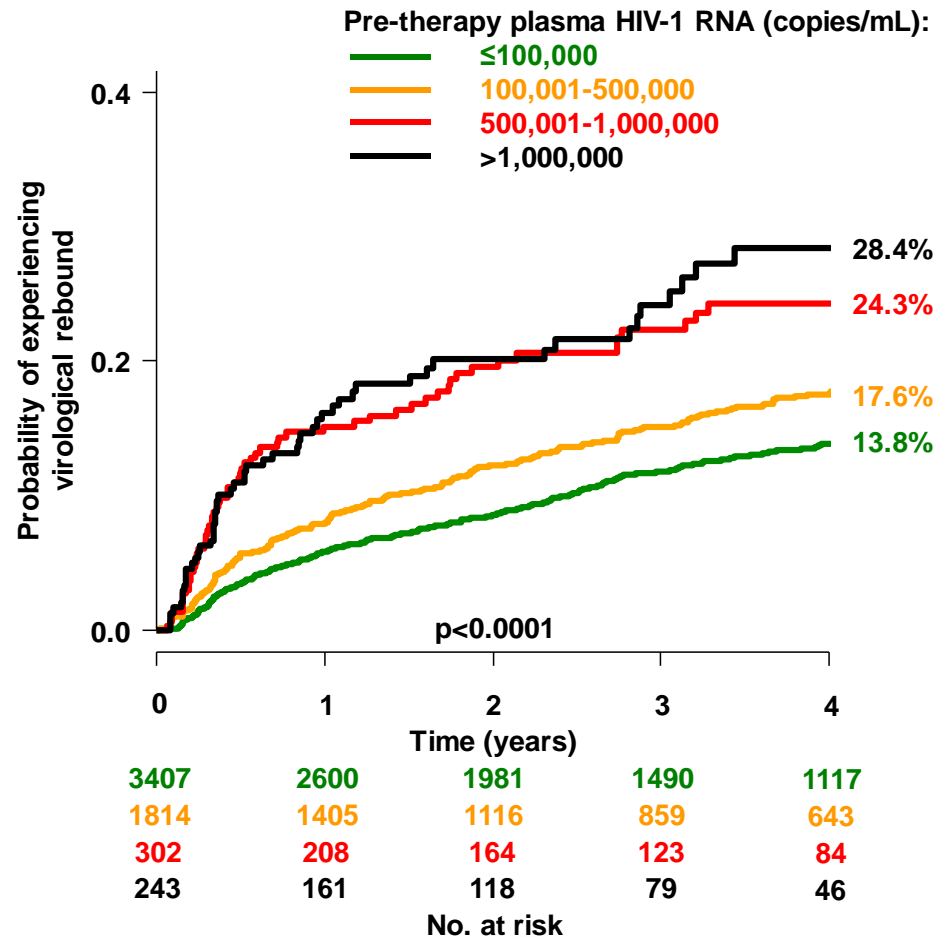
>500K



Patients (N=1,734) followed after HAART starting regardless therapy changes or interruptions. CI: confidence interval. HAART: highly active antiretroviral therapy. <30K: <30,000. 30-100K: 30,000-100,000. 100-300K: 100,000-300,000. 300-500K: 300,000-500,000. >500K: >500,000. VS: virological success.

**Pre-HAART viremia has an impact on the
achievement of virological success.....
And also on its maintenance!**

In patients starting a first-line regimen, high pre-cART viremia correlates with high probability of experiencing virological rebound after the achievement of virological success



Kaplan-Meier curves estimates of cumulative probability of virological rebound according to pre-HAART viremia ranges and type of first-line treatment

Factors influencing long-term viral suppression

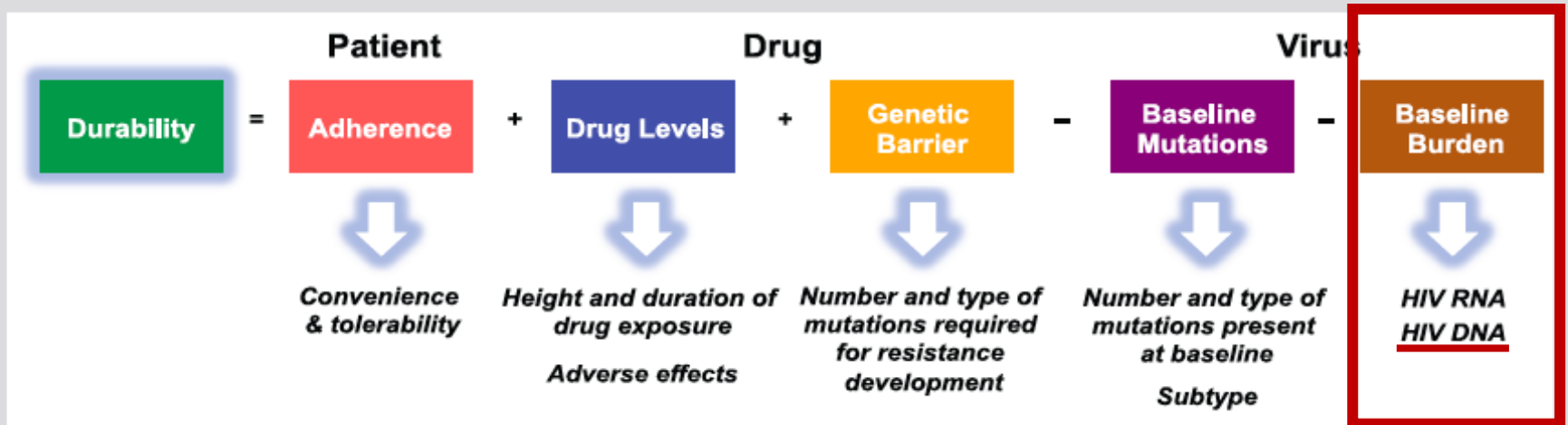
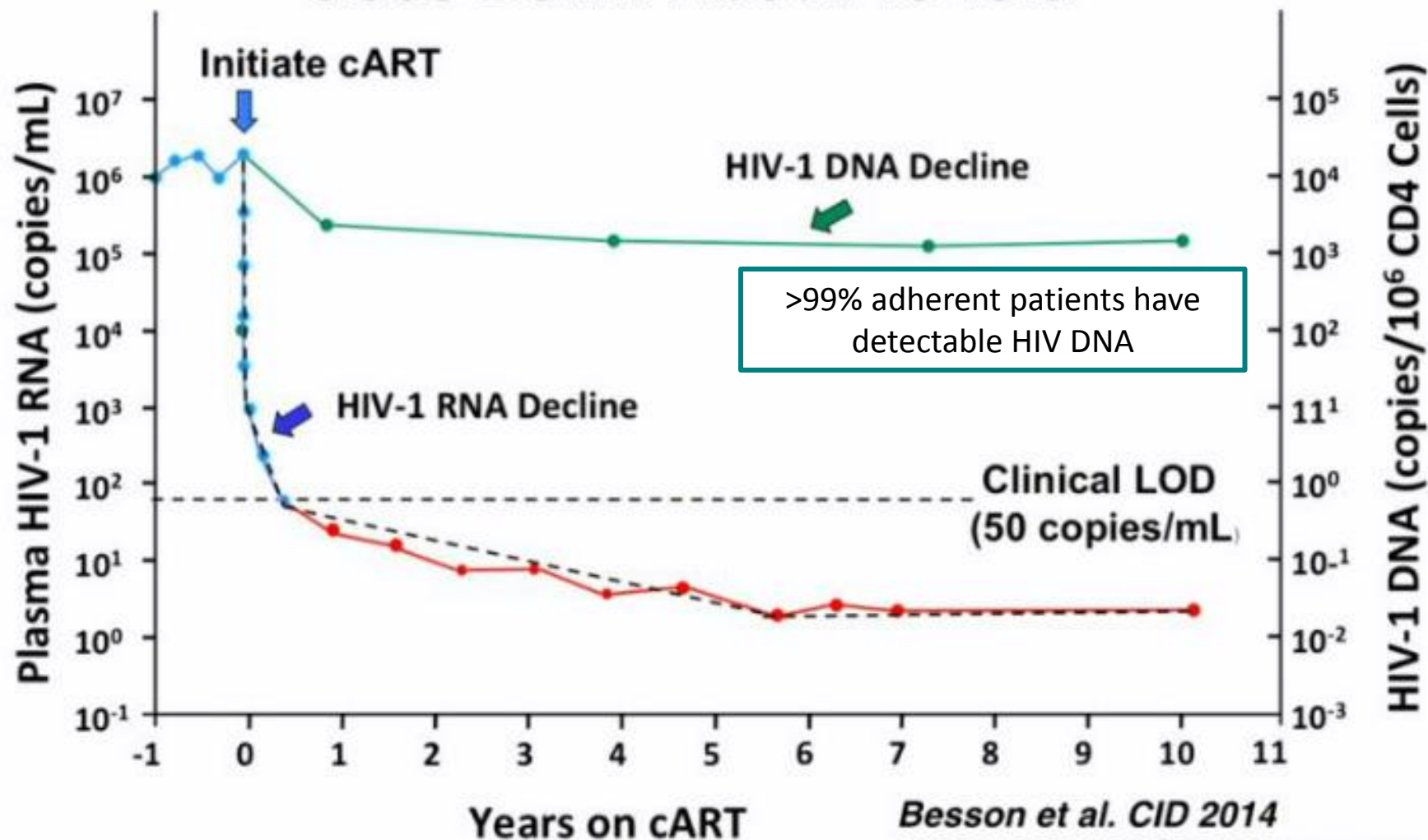


Figure 1. Factors influencing long-term viral suppression, adapted from Massimo Andreoni personal communication.

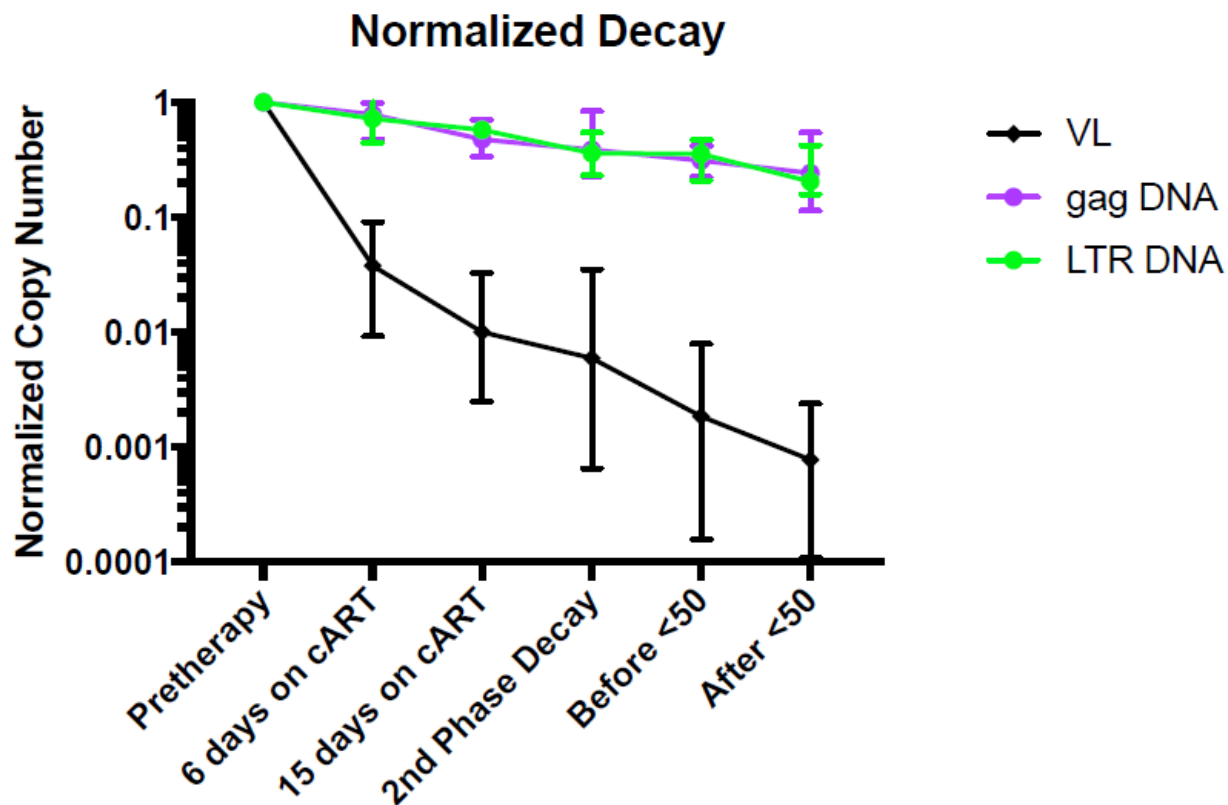
HIV-DNA load correlates directly with the number of latently HIV-infected cells that constitute the viral reservoir.

Therefore, the quantification of total HIV-DNA in PBMCs provides a reliable and easy way of measuring the size of the cellular reservoirs of HIV.

Although HIV DNA Declines on Therapy, It Only Goes Down About 15-fold



Besson et al. CID 2014
Hong et al. AIDS Reviews 2015



HIV DNA and Viral Load Suppression

All patients had successful suppression of HIV RNA to <50cps/mL plasma by a median of 139.5 days on cART.

Overall, total HIV DNA copies/million CD4+ cells decreased an average of 4.5-fold from pre-therapy to viral suppression (75%).

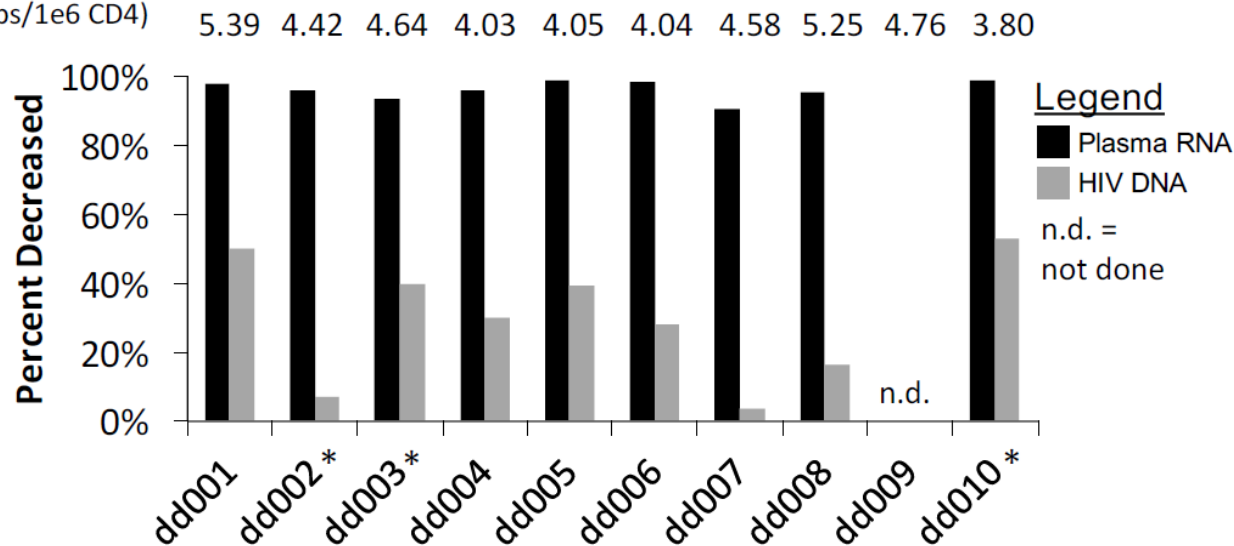
HIV Viremia is the Product of a Small Fraction of HIV Infected Cells

Elizabeth M. Anderson¹, S. Hill¹, J. Bell³, C. Rehm², S. Jones³, R. Gorelick³, M. Polis⁴, M.F. Kearney¹, J.M. Coffin⁵, and F. Maldarelli¹

¹HIV Dynamics and Replication Program, NCI, Frederick, MD, ²Laboratory of Immunoregulation, NIAID, Bethesda MD,

³Leidos Biomedical Research Inc. Frederick, MD, ⁴Collaborative Clinical Research Branch, NIAID, Bethesda MD, ⁵Department of Molecular Biology and Microbiology, Tufts University, Boston MA

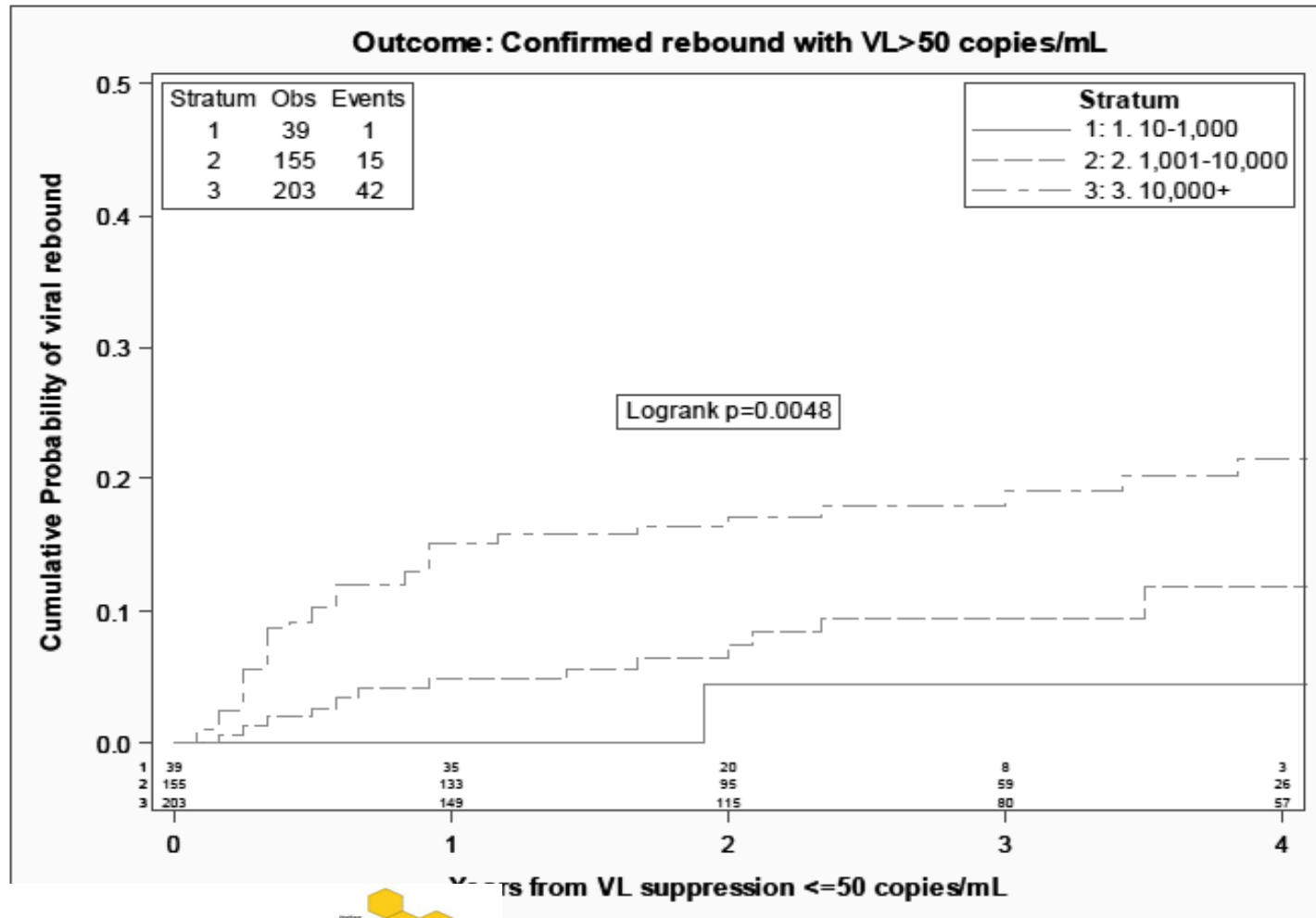
Baseline HIV LTR
DNA (Log10
cps/1e6 CD4)



Small Fraction of HIV DNA+ PBMCs is Lost after 6 days

Strikingly, while HIV RNA in the blood declined on average 96% after 6 days on therapy, HIV DNA in the blood declined an average of only 30% (mean 491cps HIV DNA/ml; range 49-903cps/ml) indicating that the majority of viremia in the blood is produced by a small fraction of HIV infected PBMCs, suggesting that each infected cell produces a median of 104cps HIV RNA in plasma (range 21.2-8999cps/cell).

Higher levels of BL HIV DNA showed an increased probability of virological rebound after virological success



Various studies have shown that the level of baseline HIV-DNA can influence the maintenance of virological success under simplification therapy

[J Med Virol](#). 2007 Jul;79(7):880-6.

Cellular HIV-1 DNA quantitation in patients during simplification therapy with protease inhibitor-sparing regimens.

[Sarmati L¹](#), [Parisi SG](#), [Nicastri E](#), [d'Ettorre G](#), [Andreoni C](#), [Dori L](#), [Gatti F](#), [Montano M](#), [Buonomini AR](#), [Boldrin C](#), [Palù G](#), [Vullo V](#), [Andreoni M](#).

[J Antimicrob Chemother](#). 2010 May;65(5):1005-7. doi: 10.1093/jac/dkq084. Epub 2010 Mar 18.

Impact of 48 week lopinavir/ritonavir monotherapy on blood cell-associated HIV-1-DNA in the MONARK trial.

[Avettand-Fenoel V¹](#), [Flandre P](#), [Chaix ML](#), [Ghosn J](#), [Delaugerre C](#), [Raffi F](#), [Ngovan P](#), [Cohen-Codar I](#), [Delfraissy JF](#), [Rouzioux C](#); MONARK Study Group.

[HIV Clin Trials](#). 2013 May-Jun;14(3):120-6. doi: 10.1310/hct1403-120.

Long-term HIV-1 virologic control in patients on a dual NRTI regimen.

[Prazuck T¹](#), [Zucman D](#), [Avettand-Fènoël V](#), [Ducasse E](#), [Bornarel D](#), [Mille C](#), [Rouzioux C](#), [Hocqueloux L](#).

Role of Baseline HIV-1 DNA Level in Highly-Experienced Patients Receiving Raltegravir, Etravirine and Darunavir/Ritonavir Regimen (ANRS139 TRIO Trial)

[Charlotte Charpentier^{1*}](#), [Catherine Fagard^{2,3}](#), [Céline Colin^{2,3}](#), [Christine Katlama⁴](#), [Jean-Michel Molina⁵](#), [Christine Jacomet⁶](#), [Benoit Visseaux¹](#), [Anne-Marie Taburet⁷](#), [Françoise Brun-Vézinet¹](#),



2013

Virological Factors Associated With Outcome of Dual ETR/RAL Therapy (ANRS-163 Trial)

[Cathia Soulie](#), [Lambert Assoumou](#), [Sophie Sayon](#), [Thuy Nguyen](#), [Marc-Antoine Valantin](#), [Virginie Ferre](#), [Chakib Alloui](#), [Brigitte Montes](#), [Véronique Avettand-Fenoel](#), [Constance Delaugerre](#), [Diane Descamps](#), [Esteban Martinez](#), [Jacques Reynes](#), [Gilles Peytavin](#), [Dominique Costagliola](#), [Christine Katlama](#), [Vincent Calvez](#), [Anne-Geneviève Marcelin](#).



PS6/4

FACTORS PREDICTING VIROLOGICAL FAILURE DURING DOLUTEGRAVIR MAINTENANCE MONOTHERAPY

[Ingeborg Wijting](#), [Sofie L. Rutsaert](#), [Casper Rokx](#), [David M. Burger](#), [Elrozy Andrinopoulou¹](#), [Linus Vandekerckhove](#), [Bart Rijnders](#)



Are we ready to test HIV DNA in clinical practice????

- Home-made technology available
- Commercial assays developed by small companies available
- Commercial assays under development by large companies as an adaptation of HIV RNA assays

Droplet Digital PCR helps improve accuracy of HIV-DNA quantification

38 Papers published from 2013 to 2016 used quantification of HIV-DNA : ddPCR ->24/38

REVIEW

Journal of Virus Eradication 2016; 2: 162–169

Diagnostic utility of droplet digital PCR for HIV reservoir quantification

Wim Trypsteen, Maja Kiselinova, Linos Vandekerckhove* and Ward De Spiegelaere

HIV Translational Research Unit, Department of Internal Medicine, Ghent University, Belgium

Digital PCR has recently been proposed as an alternative to real-time with potentially improved accuracy and precision.

In this method, each sample is divided into thousands of independent microscopic reactions prior to PCR amplification. Digital PCR provides an increased precision of four-fold to over 20-fold versus real-time, using identical quantities of clinical samples from peripheral blood.

Factors influencing long-term viral suppression

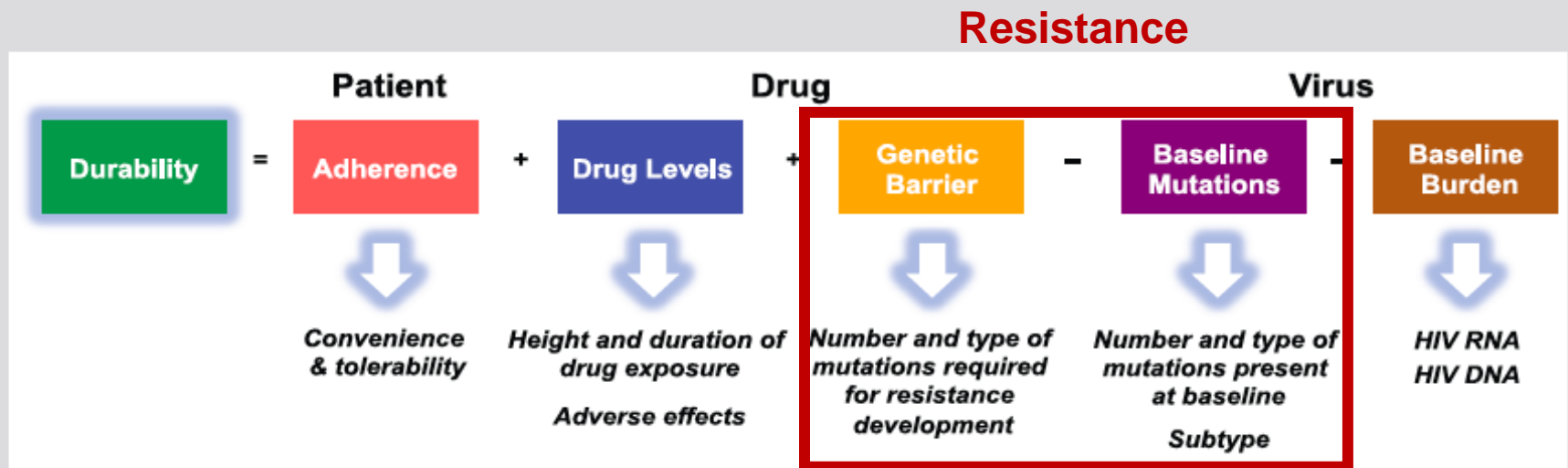
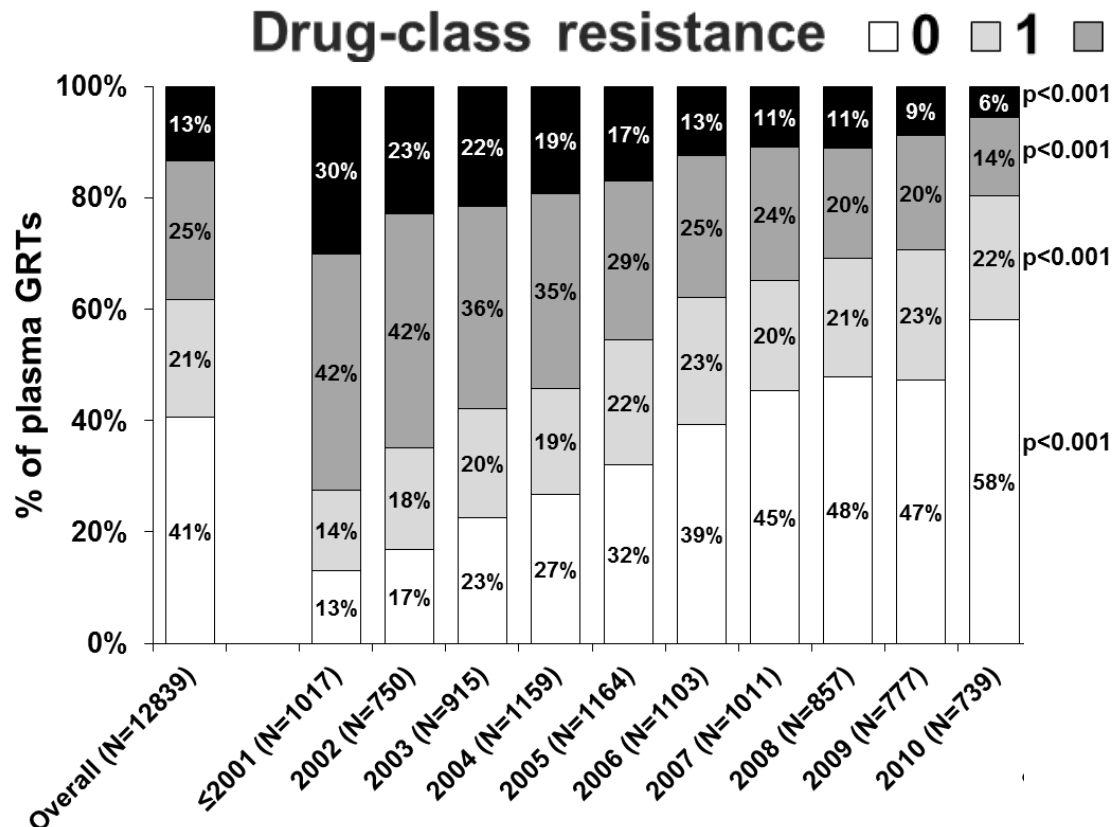


Figure 1. Factors influencing long-term viral suppression, adapted from Massimo Andreoni personal communication.

Resistance at failure significantly decreased from 1999 to 2010 in conjunction with a remarkable increase of failures without resistance.

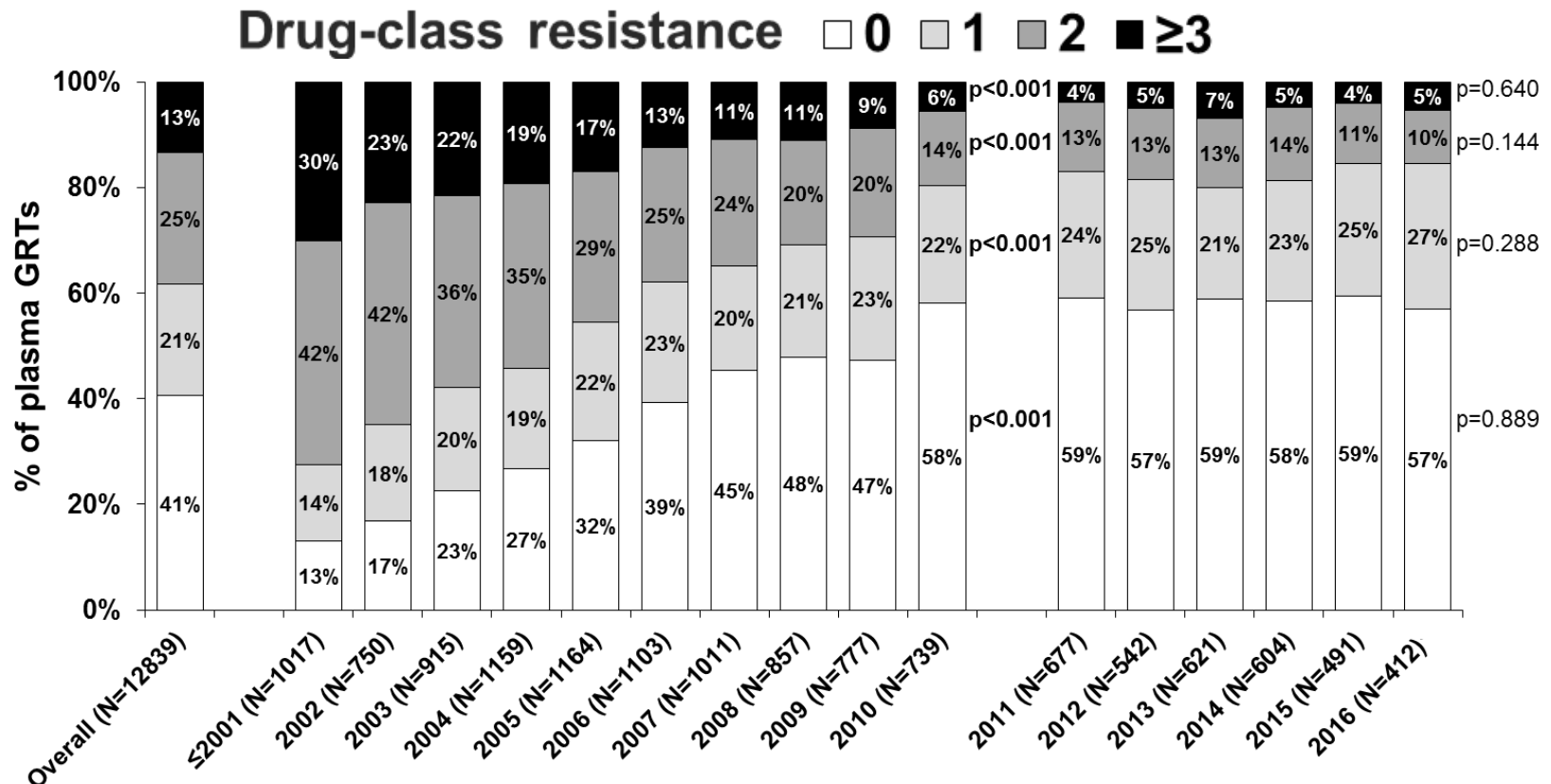
Prevalence of resistance to any drug-class among ART-experienced HIV-1 infected patients with virologic failure over the years.



Analysis performed on 14497 sequences of protease/reverse transcriptase or integrase, from 12839 GRTs performed for routine clinical practice in ART-experienced HIV-1 infected patients (N=6147). P-values were calculated by Chi-squared test for trend; statistically significant tests ($p<0.05$) are indicated in boldface. Sequences performed from 1999 to 2001 were grouped.

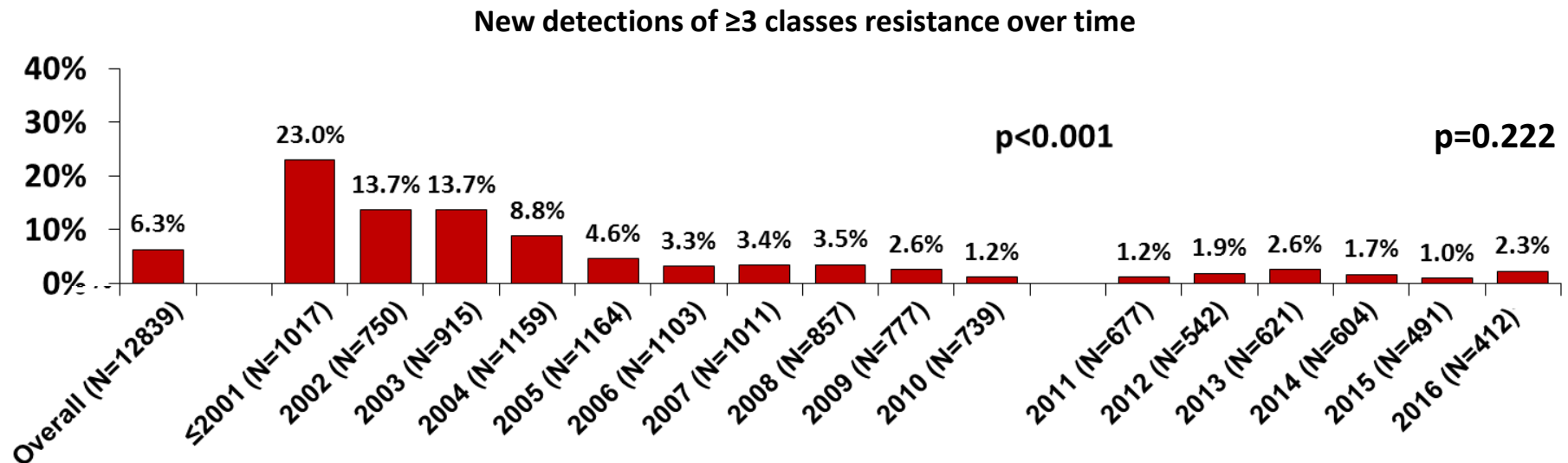
Beyond 2010, prevalence of resistance remained stable from 2011 to 2016.

Prevalence of resistance to any drug-class among ART-experienced HIV-1 infected patients with virologic failure over the years.

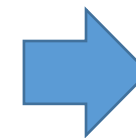
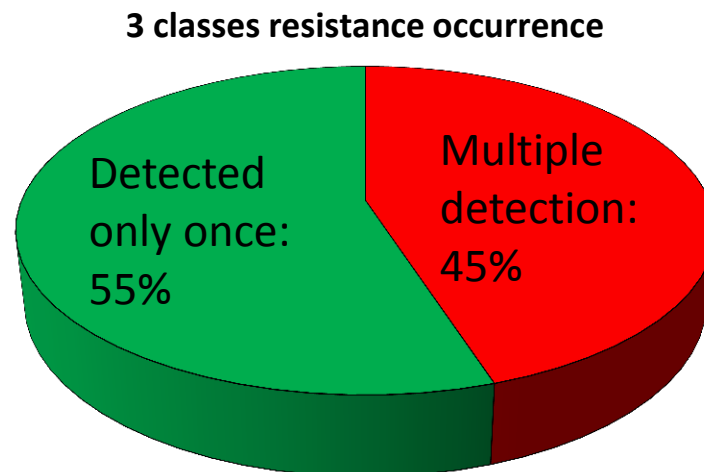
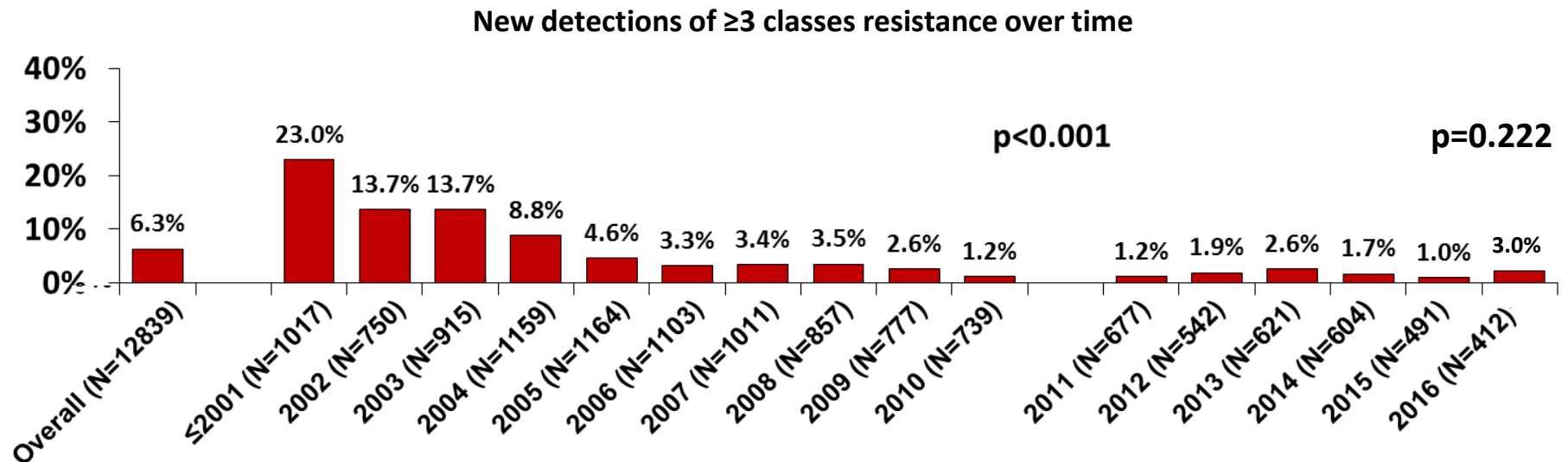


Analysis performed on 14497 sequences of protease/reverse transcriptase or integrase, from 12839 GRTs performed for routine clinical practice in ART-experienced HIV-1 infected patients (N=6147). P-values were calculated by Chi-squared test for trend; statistically significant tests ($p<0.05$) are indicated in boldface. Sequences performed from 1999 to 2001 were grouped.

New detections of ≥ 3 class-resistance dramatically decreased from 23% in 1999 to 1 % in 2010 ($p<0.001$), and remained quite stable from 2011 up to 2016 (1% to 3%, $p=0.222$).



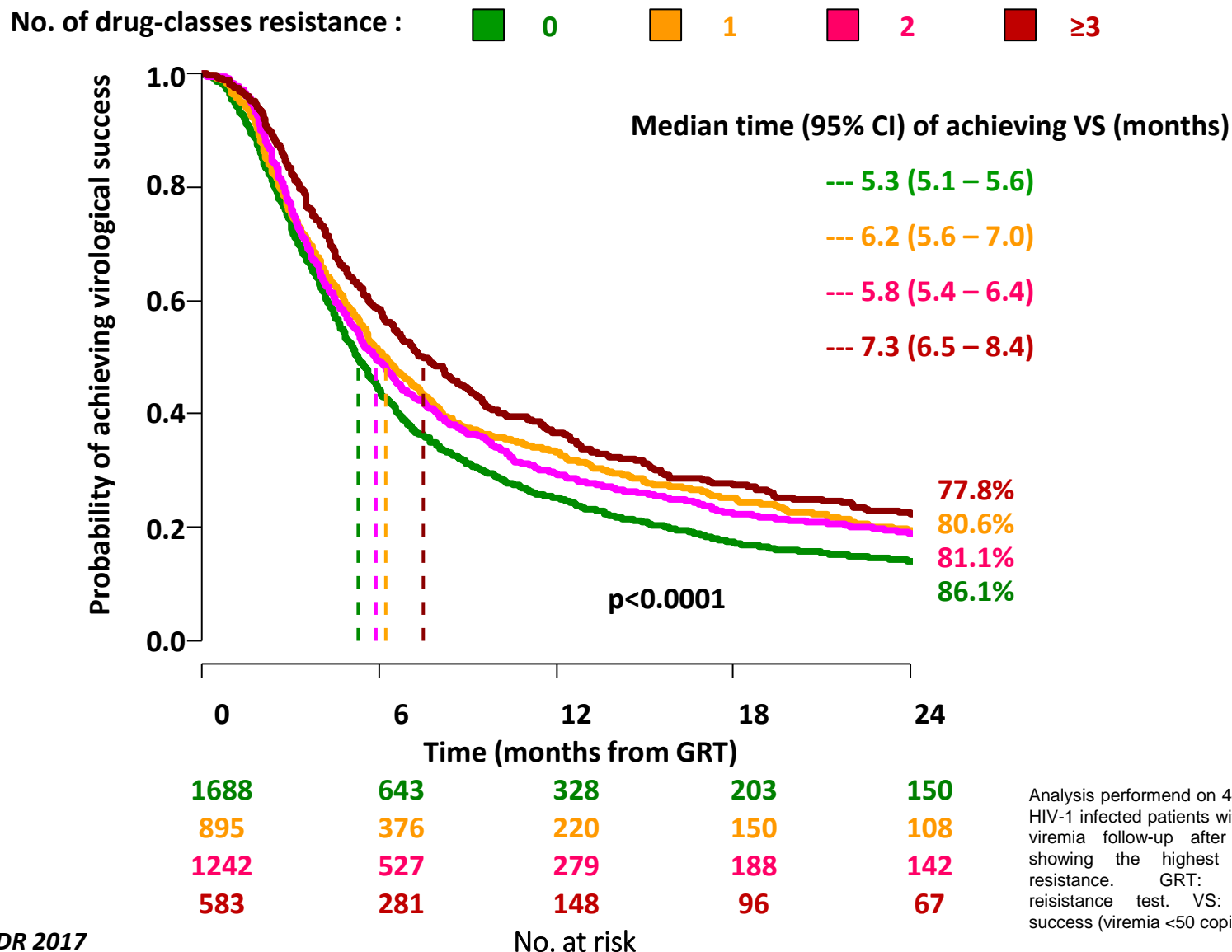
Resistance to ≥ 3 classes was found more than once in 362/810 (45%) patients, in whom re-occurred from previous GRTs recorded around 3 years before.



**Median time of
resistance
re-occurrence
3.2 (1.2-5.9) years**

- **If properly treated, also patients infected with a multi-resistant virus can achieve virological suppression**

After the latest GRT with the highest resistance class levels, the probability of achieving virological success was significantly lower in patients having 3 drug-classes resistance compared to the others.



Long-life treatment in the management of HIV infection dictates the switch!!!

Clinicians should always review possible adverse events or tolerability issues with current antiretroviral regimens.

Just because the HIV viremia is suppressed it should not be assumed that the HIV-positive person is well adapted and tolerating the current regimen.

Therefore, therapy switch is no longer a sign of therapeutic failure. If something, it represents a major option to improve chances of long-term therapeutic success

Switch Strategies for Virologically Suppressed Persons

A complete ARV history with **HIV-VL**, tolerability issues and **cumulative genotypic resistance history** should be analysed prior to any drug switch.

Pre-existent NRTI and NNRTI resistance impacts on maintenance of virological suppression in HIV-1-infected patients who switch to a tenofovir/emtricitabine/rilpivirine single-tablet regimen

D. Armenia^{1†}, D. Di Carlo^{1†}, A. Calcagno², G. Vendemiati², F. Forbici³, A. Bertoli¹, G. Berno³, S. Carta³, F. Continenza³, V. Fedele³, R. Bellagamba⁴, S. Cicalini⁴, A. Ammassari⁴, R. Libertone⁴, M. Zaccarelli⁴, V. Ghisetti², M. Andreoni⁵, F. Ceccherini-Silberstein¹, S. Bonora², G. Di Perri², A. Antinori⁴, C. F. Perno³ and M. M. Santoro^{1*}

¹Experimental Medicine and Surgery, University of Rome Tor Vergata, Rome, Italy; ²Division of Infectious Diseases, University of Turin, Turin, Italy; ³Antiretroviral Drug Monitoring Laboratory, National Institute for Infectious Diseases L. Spallanzani, IRCCS, Rome, Italy;

⁴Infectious Diseases Division, National Institute for Infectious Diseases L. Spallanzani, IRCCS, Rome, Italy; ⁵Systems Medicine, University of Rome Tor Vergata, Rome, Italy

*Corresponding author. E-mail: santormaria@gmail.com
†D. Armenia and D. Di Carlo equally contributed to this work.

Received 3 August 2016; returned 29 August 2016; revised 28 October 2016; accepted 1 November 2016

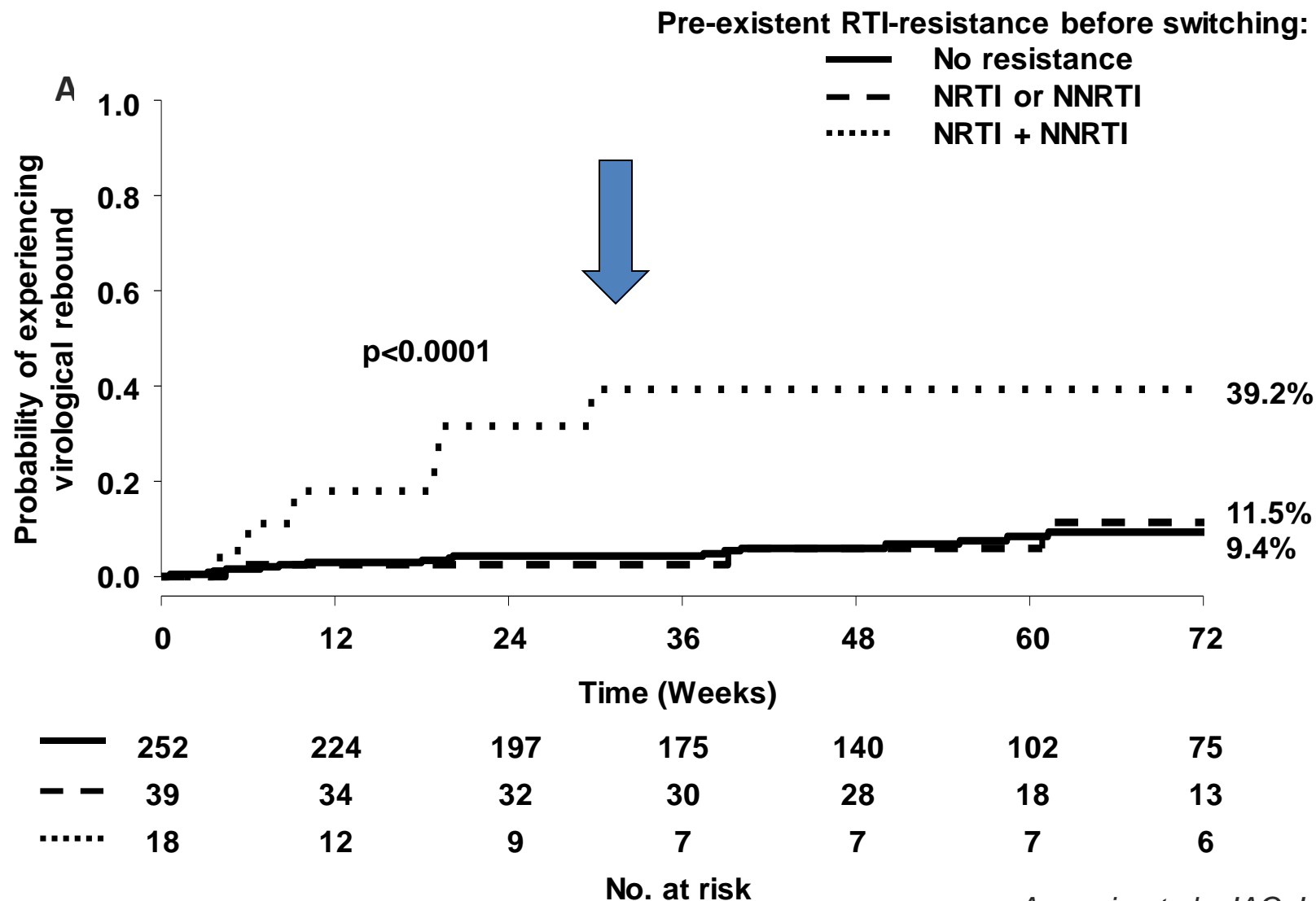
Objectives: To evaluate the maintenance of virological suppression (VS) in antiretroviral-treated HIV-1-suppressed patients switching to a tenofovir/emtricitabine/rilpivirine (TDF/FTC/RPV) single-tablet regimen, by considering pre-existent resistance (pRes).

Methods: pRes was evaluated according to resistance on all previous plasma genotypic resistance tests. Probability and predictors of virological rebound (VR) were evaluated.

Results: Three hundred and nine patients were analysed; 5.8% of them showed resistance to both NRTIs and NNRTIs, while 12.6% showed resistance to only one of these drug classes. By 72 weeks, the probability of VR was 11.3%. A higher probability of VR was found in the following groups: (i) patients with NRTI + NNRTI pRes compared with those harbouring NRTI or NNRTI pRes and with those without reverse transcriptase inhibitor pRes (39.2% versus 11.5% versus 9.4%, $P < 0.0001$); (ii) patients with a virus with full/intermediate resistance to both tenofovir/emtricitabine and rilpivirine compared with those having a virus with full/intermediate resistance to tenofovir/emtricitabine or rilpivirine and those having a virus fully susceptible to TDF/FTC/RPV (36.4% versus 17.8% versus 9.7%, $P < 0.001$); and (iii) patients with pre-therapy viraemia $>500\,000$ copies/mL compared with those with lower viraemia levels ($>500\,000$: 16.0%; 100 000–500 000: 9.3%; $<100\,000$ copies/mL: 4.8%, $P = 0.009$). pRes and pre-therapy viraemia $>500\,000$ copies/mL were independent predictors of VR by multivariable Cox regression.

Conclusions: TDF/FTC/RPV as a treatment simplification strategy shows a very high rate of VS maintenance. The presence of pRes to both NRTIs and NNRTIs and a pre-therapy viraemia $>500\,000$ copies/mL are associated with an increased risk of VR, highlighting the need for an accurate selection of patients before simplification.

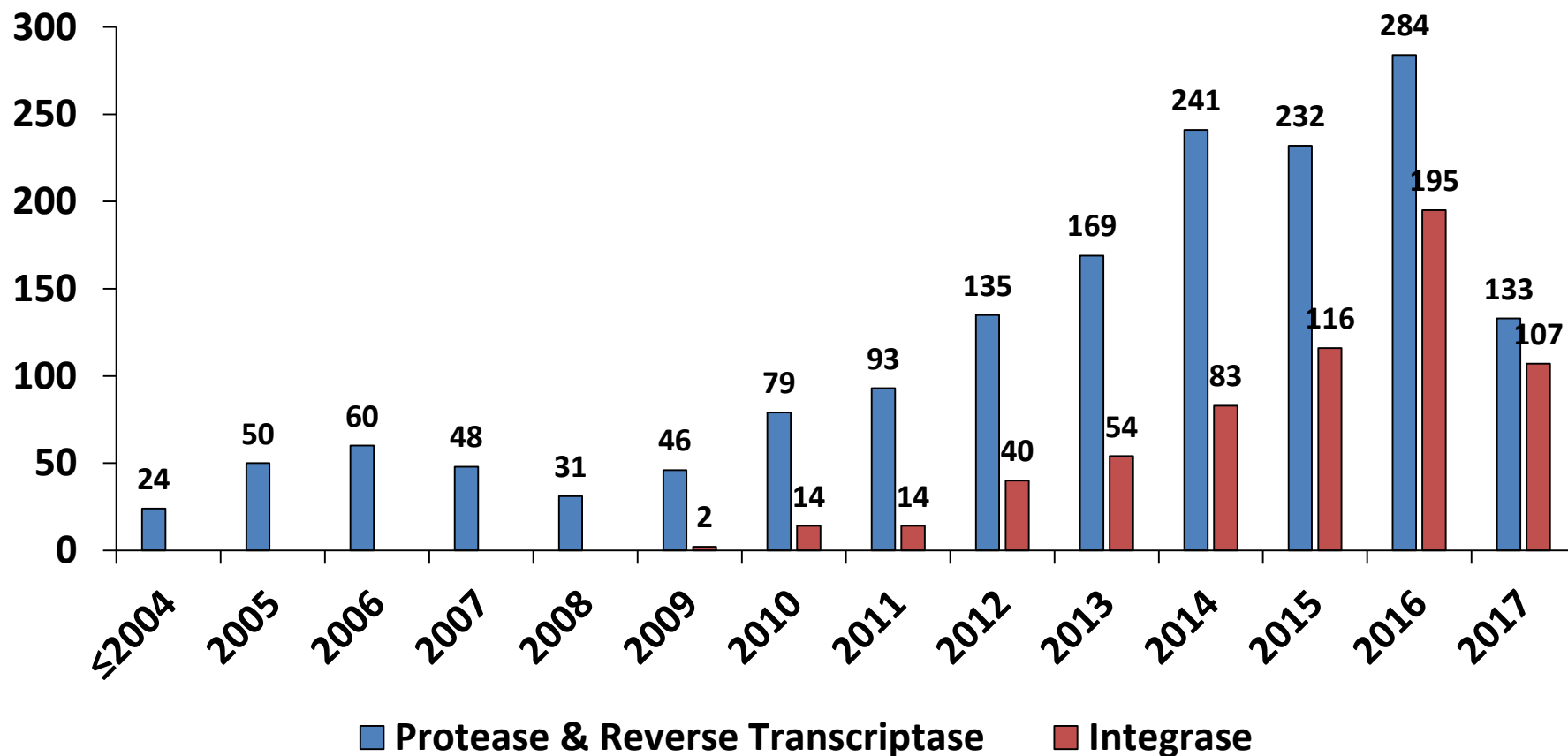
Patients with pre-existent NRTI+NNRTI resistance had a higher probability of experiencing VR compared to those harboring pre-existent NRTI or NNRTI resistance and to those without pre-existent RTI resistance.



.....As well as the genotypic test on HIV-DNA

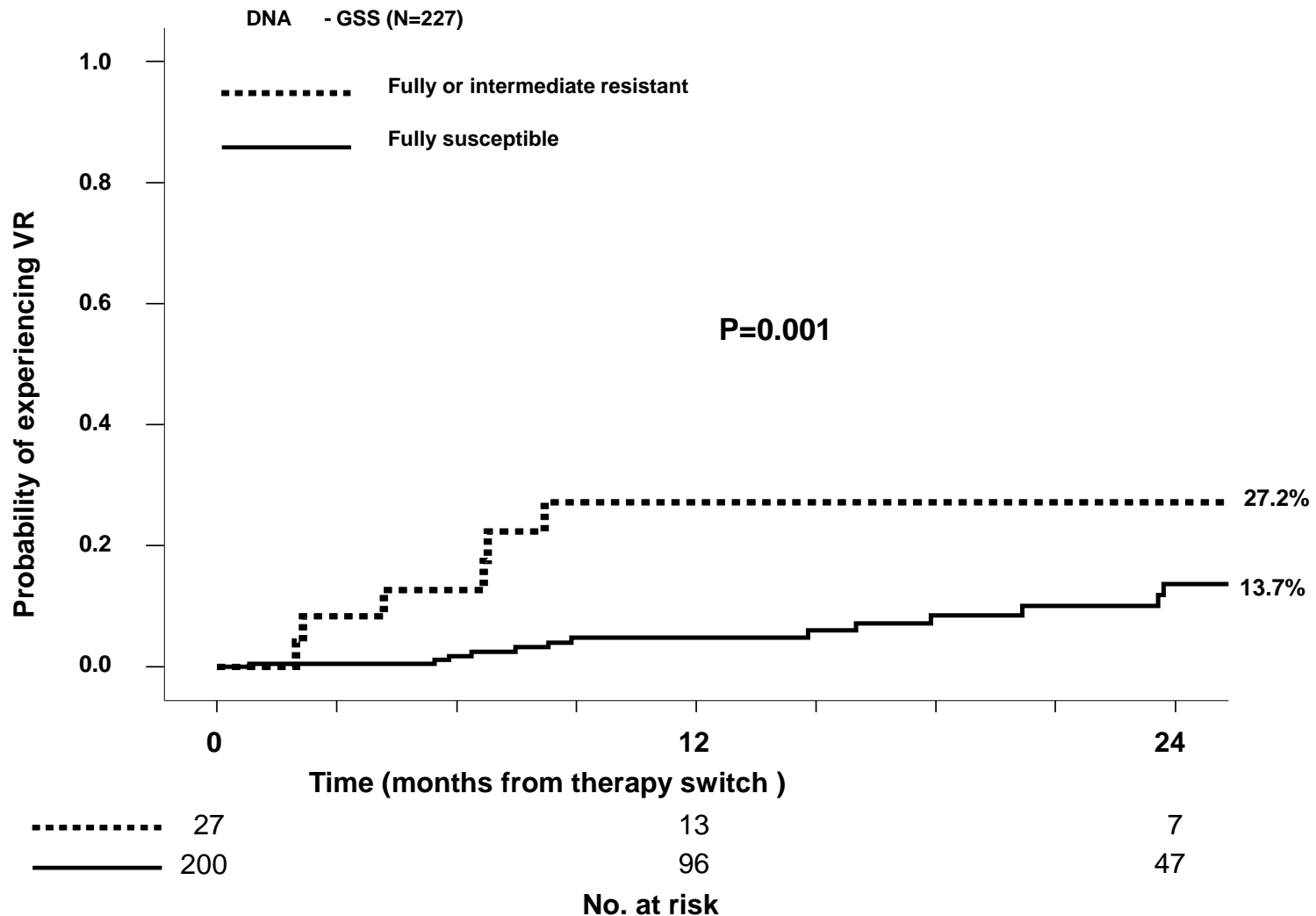
Increased requests of HIV DNA GRT in clinical practice over the recent years

Number of PBMCs genotypic resistance tests performed



Armenia D unpublished data, data updated in June 2017

Virologically suppressed patients showing an intermediate or fully resistant virus at PMBC GRTs performed before a therapy switch had a higher probability of experiencing virological rebound compared to those carrying a fully susceptible virus



HIV DNA Genotypic Resistance Test

is a good tool for therapy optimization

in both drug-naïve and drug-experienced patients

1. Sarmati L, Nicastri E, Uccella I, et al. 2003. J Clin Microbiol **41**:1760-2.
2. Parisi SG, Boldrin C, Cruciani M, et al. 2007. J Clin Microbiol **45**:1783-1788.
3. Turriziani O, Bucci M, Stano A, et al. 2007. J Acquir Immune Defic Syndr **44**:518-524.
4. Palmisano L, Galluzzo CM, Giuliano M. 2009. J Acquir Immune Defic Syndr **51**:233-234.
5. Banks L, Gholamin S, White E, et al. 2012. J AIDS Clin Res **3**:141-147.
6. Delaugerre C, Braun J, Charreau I, et al. 2012. HIV Med **13**:517-525.
7. Bon I, Turriziani O, Musumeci G, et al. J Med Virol 2015 **87**:315-322.
8. Fabeni L, Berno G, Svicher V, et al. 2015. J Clin Microbiol **53**:2935-41.
9. Gallien S, Charreau I, Nere ML, et al. 2015. J Antimicrob Chemother **70**:562-565.
10. Lubke N, Di Cristanziano V, Sierra S, et al. 2015. Intervirology **58**:184-189.
11. Gantner P, Morand-Joubert L, Sueur C, et al. 2016. J Antimicrob Chemother **71**:751-61.
12. Michelini Z, Galluzzo CM, Pirillo MF, et al. 2016. J Med Virol. doi: 10.1002/jmv.24581.
13. Fernández-Caballero JÁ, Chueca N, Álvarez M, et al. 2016. BMC Infect Dis. **16**:197.
14. Zaccarelli M, Santoro MM, Armenia D, et al. 2016. J Clin Virol **82**:94-100.
15. Lambert-Niclot S, Allavena C, Grude M, et al. 2016. J Antimicrob Chemother. 71:2248-51.
16. Rodallec A, Le Guen L, Leplat A, et al. 2017. IAS. Abstract MOPEB0270.
17. Allavena C, Rodallec A, Leplat A, et al. J Virol Methods. 2018 Jan;251:106-110.

Conclusions

- More high sensitive and specific molecular tests are fundamental for the long term monitoring of HIV infection.
- HIV infected patients often maintain detectable virological parameters not considered in the past, such as very low-level viremia and HIV-DNA.
- These parameters deserve particular attention in preventing chronic inflammation and in reducing viral reservoir.
- Resistance testing is still today an important tool to tailor a correct therapy.
- The improvement of resistance testing, finalized to detect resistance even at undetectable viremia (on DNA), allows clinicians to optimize therapy in the case of switch for treatment simplification.



Acknowledgements



University of Milan, Milan Italy: C. F. Perno, D. Di Carlo.

University of Rome Tor Vergata, Rome Italy: F. Ceccherini Silberstein, V. Svicher, A. Bertoli, D. Armenia, C. Alteri, M.C. Bellocchi, L. Fabeni, R. Salpini, A. Biddittu, M. Romani, M. Bruni, L. Carioti, P. Saccomandi, R. Scutari.

Policlinic of Rome Tor Vergata, Rome Italy: M. Andreoni, L. Sarmati, A.R. Bonomini, L. Dori, E. Gentilotti, D. Maffongelli. A. Ricciardi, M. Viscione, S. Gini, C. Cerva, V. Malagnino, T. Guenci, F. Stazi, S. Giannella, V. Serafini, M. Montano, M. Ciotti, P. Paba. C. Favalli

INMI L Spallanzani, Rome, Italy: A. Antinori, G. D'Offizi, N. Petrosillo, U. Visco-Comandini, G. Liuzzi, F. Antonucci, E. Boumis, P. De Longis, E. Nicastrì, A. Ammassari, R. Bellagamba, M. Zaccarelli, C. Pinnetti, S. Cicalini, A. Sanpaoloesi, G. De Carli, F.M. Fusco, L. Lo Iacono, M.L. Giancola, R. Acinapura, P. Scognamiglio, N. Orchi, E. Girardi, M.R. Capobianchi, C. Gori, F. Forbici, S. Carta, V. Fedele, G. Bero, D. Pizzi, F. Continenza, R. D'Arrigo, A. Giannetti, P. Lorenzini, A. Navarra, R. Libertone, G. Ippolito.

San Gallicano Hospital, Rome, Italy: A. Latini, M. Colafigli, M. Giuliani, A. Pacifici, A. Cristaudo. **General Hospital Umberto I:** V. Vullo, G. D'Ettore, F. Falasca, O. Turriziani, G. Antonelli. **San Giovanni Addolorata Hospital, Rome, Italy:** F. Montella, F. Di Sora, W. Leti, F. Iebba. **Sant'Andrea Hospital, Sapienza University, Rome, Italy:** A. Pennica. **Rebibbia, Rome, Italy:** S. Marcellini. **Bambin Gesù Hospital, Rome Italy:** S. Bernardi. **Polo Pontino, Sapienza University, Rome, Italy:** C. Mastroianni, M. Lichtner, V.S. Mercurio, C. Del Borgo, R. Marrocco. **Frosinone Hospital, Frosinone, Italy:** G. Farinelli, E. Anzalone, M. Limodio, L. Sarracino. **Rieti Hospital, Italy:** G. Natalini Raponi, M.E. Bonaventura. **Viterbo Hospital, Viterbo, Italy:** O. Armignacco, G. Bernardini, A. Caterini, F. Ferri, A. Ialungo, E. Liguori, D. Migliorini, R. Monarca, R. Preziosi, E. Rastrelli, G. Starnini, G. Sebastiani.

University of Turin, Turin, Italy: G. Di Perri, S. Bonora, A. Calcagno, V. Ghisetti, G. Vandemmiati, T. Allice.

Modena Hospital, Modena, Italy: C. Mussini, V. Borghi, W. Gennari, A. Cossarizza, M. Nasi, M. Di Gaetano.

Pescara General Hospital, Pescara, Italy: G. Parruti, F. Vadini, F. Sozio, E. Mazzott, T. Ursini, E. Polilli, P. Di Stefano, M. Tontodonati, G. Calella. **San Salvatore, L'Aquila, Italy:** A. Grimaldi, A. Cellini. **Ancona Hospital, Ancona, Italy:** A. Mataloni Paggi. **Giuseppe Mazzini Hospital, Teramo, Italy:** Di Giammartino, L. Falconi, P. Tarquini. **San Salvatore – Muraglia- Hospital, Pesaro, Italy:** E. Petrelli, G. Corbelli, P. Tarquini. **Avezzano Hospital, Avezzano, Italy:** M. Paoloni, R. Mariani. **AO Papa Giovanni XXIII, Bergamo, Italy:** F. Maggiolo, AP Callegaro. **AO Careggi, Florence, Italy:** K. Sterrantino.

Cotugno Hospital, Naples, Italy: A. Chirianni, M. Gargiulo. **University of Campania Vanvitelli, Italy:** S. Marini, N. Coppola. **Bisceglie-Trani Hospital, Bisceglie, Italy:** R. Losappio. **Catania Hospital, Catania, Italy:** R. La Rosa. **Enna Hospital, Enna, Italy:** L. Guarneri. **Palermo Hospital, Palermo, Italy:** F. Di Lorenzo T. Prestileo.

Thank you for the attention!